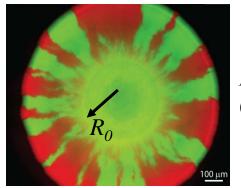
The Physics of Life: Spatial Population Genetics

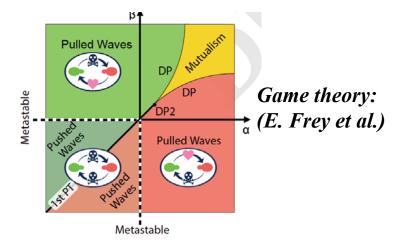
I. Introduction to spatial population genetics

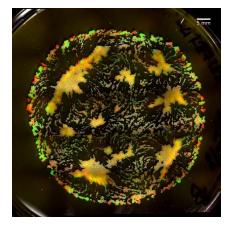
II. Pushed genetic waves and antagonistic interactions

III. Microbial interactions and expansions on liquid substrates



P. Aeruginosa (J. Xavier et al.)





S. cerevisea (S. Atis et al.)

"Clash of Genomes": Range expansions with neutrality, selection advantage, cooperation or competition

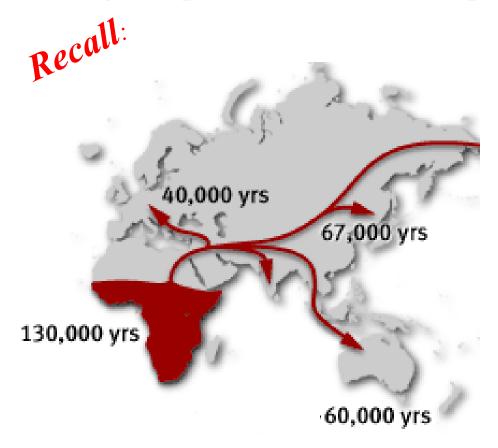
What happens at the border?



Collision of two neutral genetically labelled E. Coli colonies on hard agar....

Hernan Garcia, Rob Phillips & drn

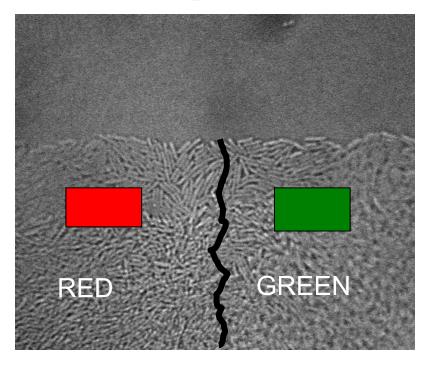
Range Expansions with Competition or Cooperation



In 500 generations....

Large mammals expand over $\sim 10^4$ km

Bacteria (in a Petri dish) expand ~ 1 cm

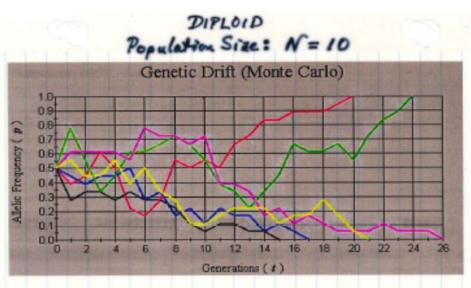


See Populus program...

Red and Green Strains...

- 1. Could be neutral
- 2. Could have different doubling times
- 3. One or both could secrete amino acids useful to the other (mutualism)
- 4. One or both could secrete toxins that impede the other (competive exclusion)

Genetic drift in well-mixed neutral models



J.F. CROW & M. Kinnera

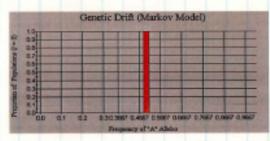
Allele frequencies diffuse due to genetic drift

M. Kimura, Genetics 47, 713 (1962)

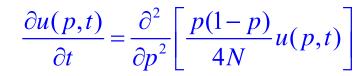
I random walk dynamics of gene frequencies!

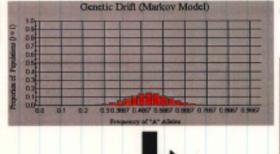
u(p,t)= probability allele A has frequency p at time t.

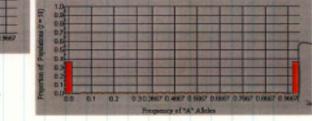
Finite populations go to fixation for long times.



* All finite populations eventually fixed with homozygous population of all (a/a) or all (NA)

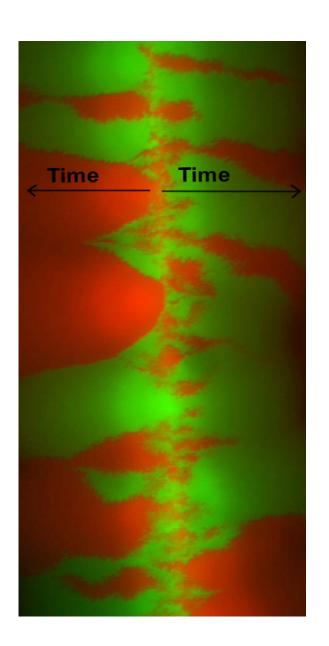


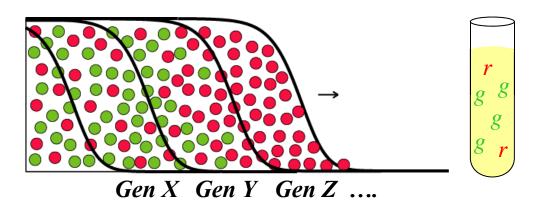




Genetic Drift (Markov Model)

Linear Inoculations: "Genetic demixing" results from number fluctuations at the frontier

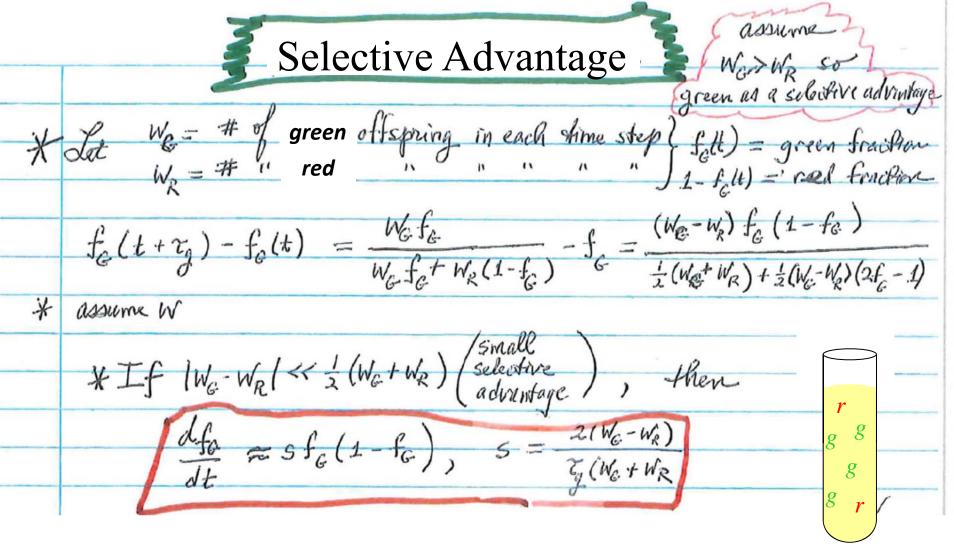




◆In effect, a moving population front is a serial dilution experiment in a well mixed test tube

How do we add a selective advantage?

go to board 1



$$f_G(t) = \frac{f_G(0)e^{st}}{1 + f_G(0)(e^{st} - 1)}$$

FKPP equation for a genetic wave

$$\frac{\partial f_G(x,t)}{\partial t} = D \frac{\partial^2 f_G}{\partial x^2} + s f_G (1 - f_G)$$

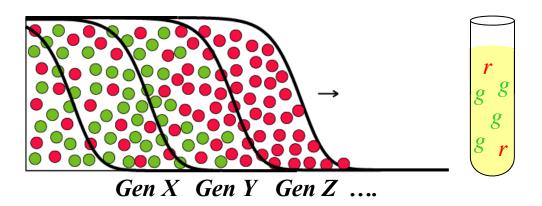
For a "zero-dimensional" frontier, f(t), the fraction of red cells with selective advantage s at time t obeys

$$\frac{df(t)}{dt} = sf(1-f) + \sqrt{\frac{f(1-f)}{2N}} \Gamma(t)$$

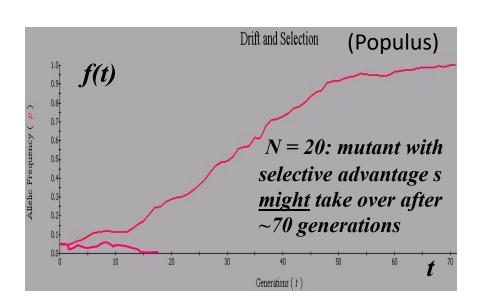
$$< \Gamma(t)\Gamma(t') >= \delta(t-t') \text{ (Ito calculus)}$$

$$s = 2\frac{w_R - w_G}{w_R + w_G}$$

Genetic Drift & Selection

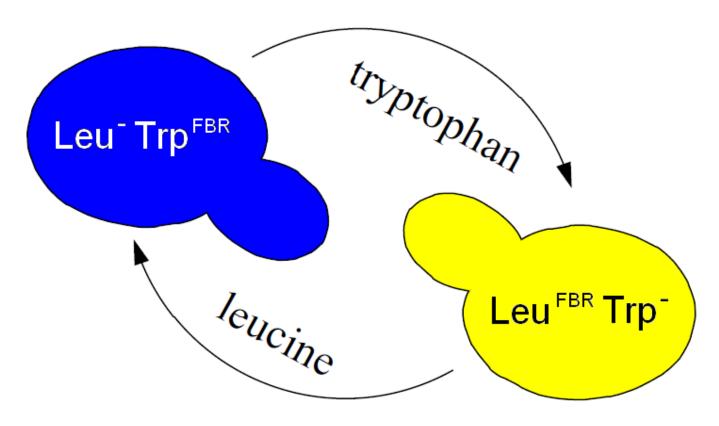


◆In effect, a moving population front is a serial dilution experiment in a well mixed test tube



Range expansions for mutualists

M. Mueller, A. Murray K. Korolev & drn



FBR = "feed back resistant"

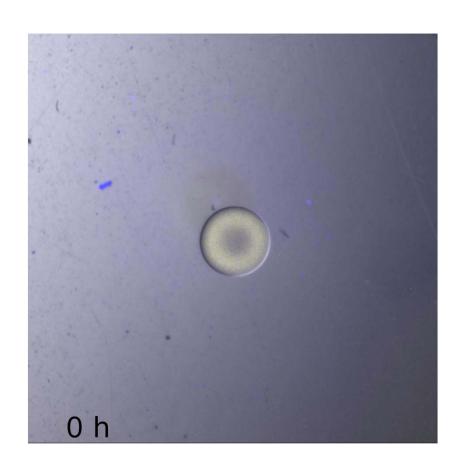
Mutualism: survival requires exchanging amino acids, if leucine and tryptophan are not already present in the natural environment

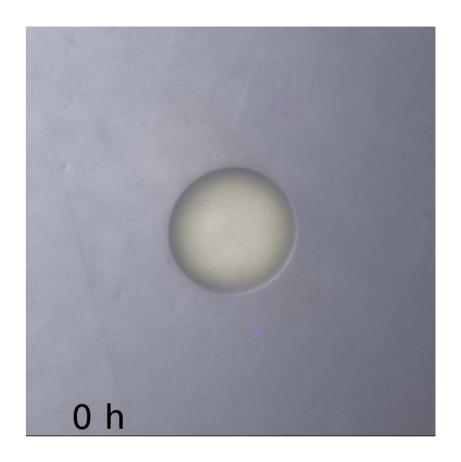
Mutualists on various substrates (Mueller, Murray, drn)

CSM (abundant Leu & Trp)

→ Mutualism unimportant

CSM-Leu-Trp (Leu, Trp missing)
→ Obligate mutualism



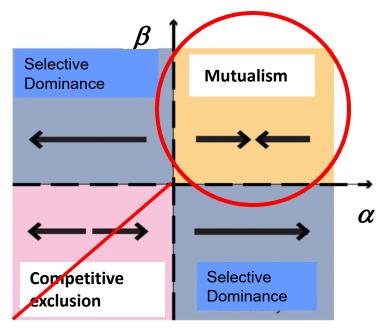


go to board 3

Game Theory and Frequency-Dependent Selection

If w_B and w_Y are the number of blue and yellow offspring produced during one generation at a given point on the frontier...

selective advantage $s \approx 2 \frac{w_B - w_Y}{w_B + w_Y}$



M. Nowak et al., Nature (2004)

J. Gore et al. Nature (2009)

E. Frey et al., Phys. Rev. Lett. (2010)

If f(x,t) is the yellow fraction, describe mutualism by... $s(f) \approx s_0(f^* - f)$ $w_Y(x,t) = g + \beta(1 - f(x,t))$ $s_0 = (\alpha + \beta)/g$ $w_B(x,t) = g + \alpha f(x,t)$ $f^* = \beta/(\alpha + \beta)$

$$\frac{\partial f(x,t)}{\partial t} = D \frac{\partial^2 f(x,t)}{\partial x^2} + s_0(f^* - f)f(1 - f)$$

 $\alpha, \beta > 0 \rightarrow \text{Mutualism}$

frequency dependent selection:

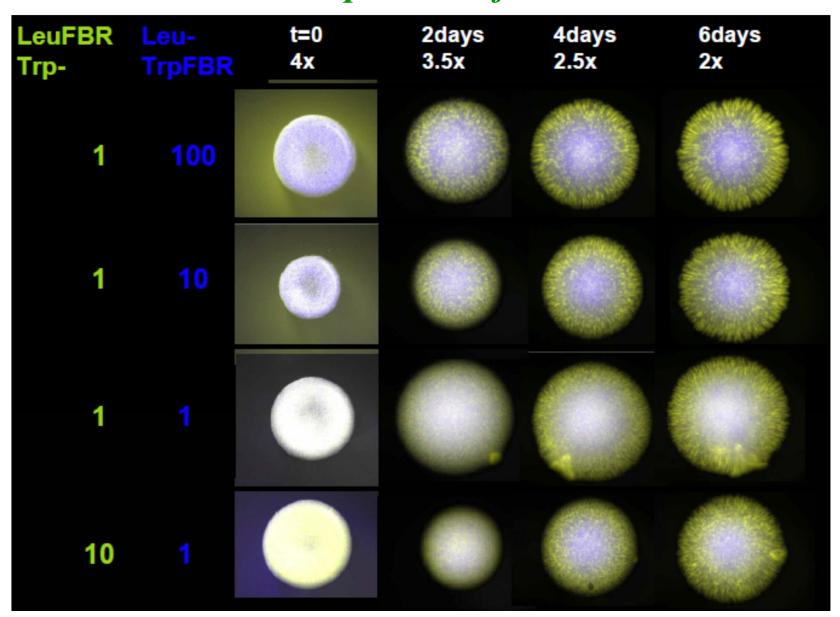
$$S_0(f^* - f)f(1 - f)$$

$$f^*$$

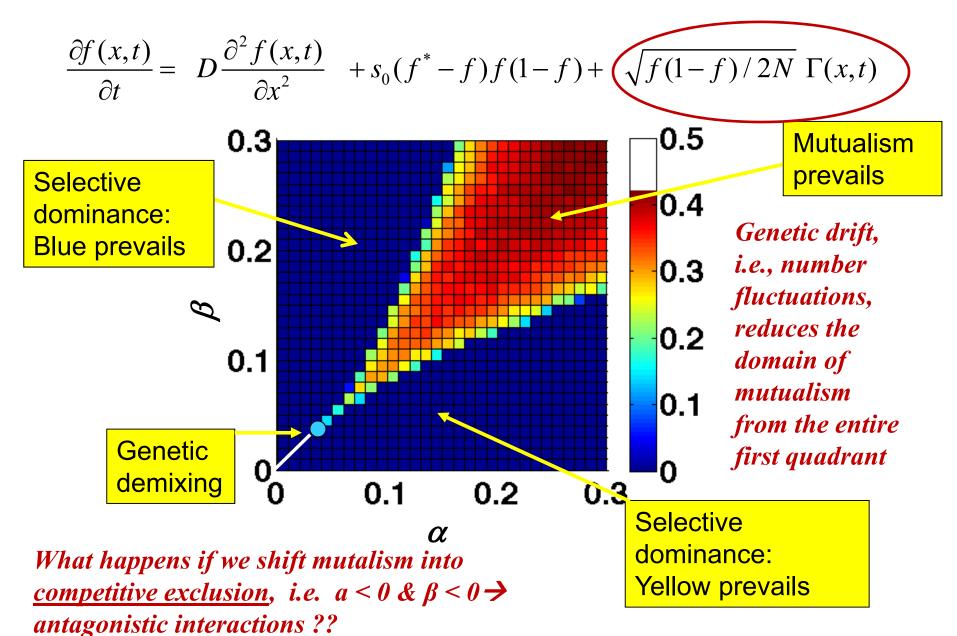
$$0$$

$$1$$

Experiments reveal that an "evolutionary stable strategy" is indeed reached independent of the initial condition...



Effect of genetic drift for $\alpha > 0$, $\beta > 0$



Pushed Genetic Waves and Antagonistic Interactions

- For pushed genetic waves, a "critical nucleus" is required to excite the wave to get it started (like nucleation theory)
- How can an excitable pushed wave from a gene drive be stopped? (They are fragile and can be stopped by obstacles....)
- Can we detect the existence of a critical nucleus with killer yeast or bacteria strains? (project with Andrea & Andrew)



Andrea Giometto

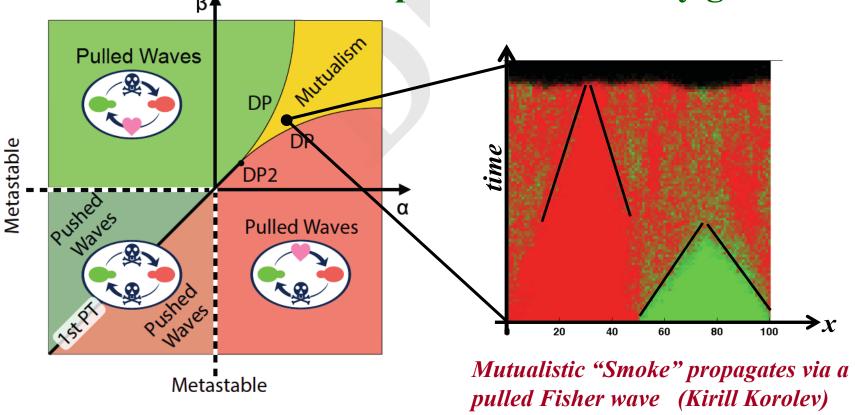


Andrew Murray



Max Lavrentovich Univ. of Tennessee

Summary: Pushed waves appear in competitive exclusion models for spatial evolutionary games



- Fisher genetic waves are "pulled waves", driven by growth and diffusive dispersal at the leading edge.
- Excitable genetic waves are "pushed waves", driven by populations behind the front whose offspring spill over to the leading edge \rightarrow strong "Allee effect" (A. Stokes, 1976)

Frequency dependent selection

If w_B and w_Y are the reproduction rates of blue and yellow produced during one generation at a given point on the frontier...

selective advantage
$$s(f) \approx \frac{w_B - w_Y}{(w_R + w_Y)/2} = generation time$$

$$\frac{\partial f(x,t)}{\partial t} = D$$

If f(x,t) = yellow fraction, 1 - f(x,t) = blue fraction describe mutualism by...

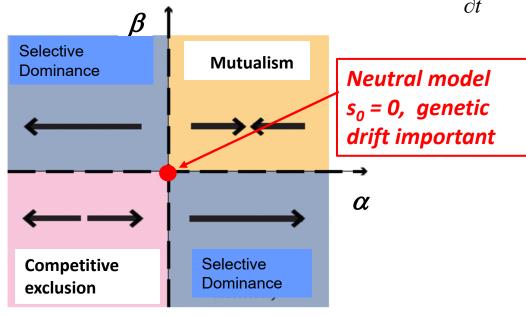
$$w_{Y}(x,t) = [1 + \beta(1 - f(x,t))] / \tau_{g} \qquad s(f) \approx s_{0}(f^{*} - f)$$

$$w_{B}(x,t) = [1 + \alpha f(x,t)] / \tau_{g} \qquad s_{0} = \alpha + \beta$$

$$\tau = \text{generation time} \qquad f^{*} = \beta / (\alpha + \beta)$$

 $\frac{\partial f(x,t)}{\partial t} = D \frac{\partial^2 f(x,t)}{\partial x^2} + s_0(f^* - f)f(1 - f)$

+ number fluctuations



M. Nowak et al., Nature (2004)

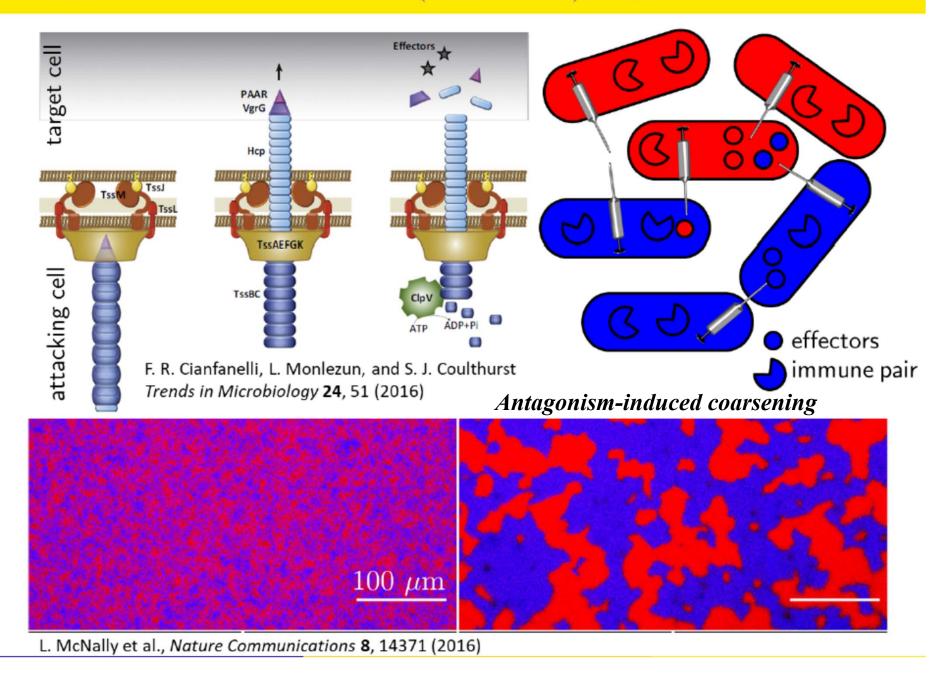
J. Gore et al. Nature (2009)

E. Frey et al., Phys. Rev. Lett. (2010)



What happens for both $\alpha \& \beta$ negative?

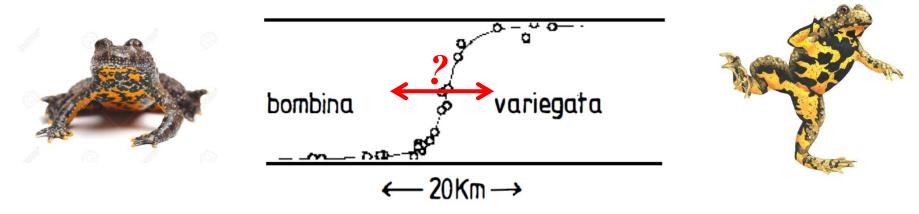
Antagonistic interactions $(\alpha, \beta < 0)$: type VI secretion



Pushed genetic waves in "hybrid zones"

N. H. Barton & G. M. Hewitt Ann. Rev. Ecol. & Sys. **16**, 113 (1985)

➤ Hybrid zones are narrow regions in which genetically distinct populations meet, mate and produce hybrids. Hundreds of examples known. (e.g., the grasshopper *Podisma pedestris*, the butterfly *Heliconis*.) Hybrid zones can be a few hundred meters thick and hundreds of kilometers long



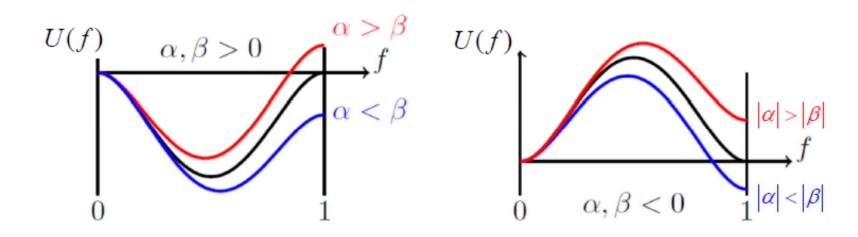
Inferred profile from electrophoretic variations across the hybrid zone of the toads *Bombina bombina* and *Bombina variegata* near Cracow, Poland

- ➤ Which way the interface moves depends on more than just the selective advantage for example, recombination near the interface can break up favorable clusters of genes.
- In some cases, boundaries can exhibit a kind of surface tension, as well as a pressure to advance in a particular direction. This may promote sympatric speciation

A course-grained description: diffusion in a potential

Consider an allele fraction $f \equiv f(\mathbf{x}, t)$ at position \mathbf{x} and time t. Then:

$$\begin{split} \partial_t f &= D_s \nabla_{\mathbf{x}}^2 f + \frac{f(1-f)}{\tau_g} \left[(\alpha + \beta) \left(\frac{1}{2} - f \right) + \frac{\alpha - \beta}{2} \right] + \sqrt{\frac{2f(1-f)}{N\tau_g}} \, \xi \\ &= D_s \nabla_{\mathbf{x}}^2 f - \frac{1}{\tau_g} \frac{dU(f)}{df} + \sqrt{D_g f(1-f)} \, \xi \quad \text{with number fluctuations:} \\ \langle \xi(\mathbf{x},t) \rangle &= 0 \quad \text{and} \quad \langle \xi(\mathbf{x},t) \xi(\mathbf{x}',t') \rangle = \delta(t-t') \delta(\mathbf{x}-\mathbf{x}'). \end{split}$$

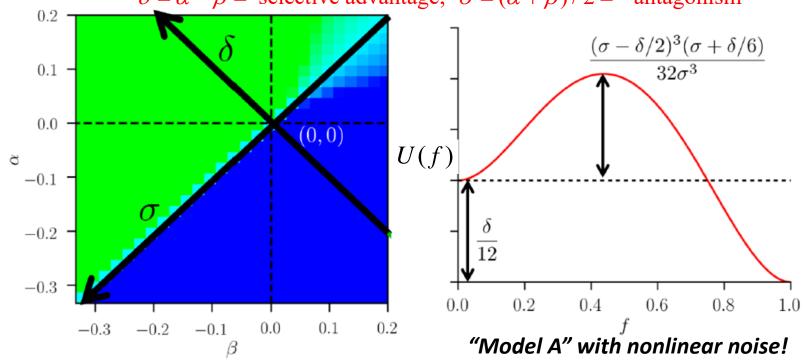


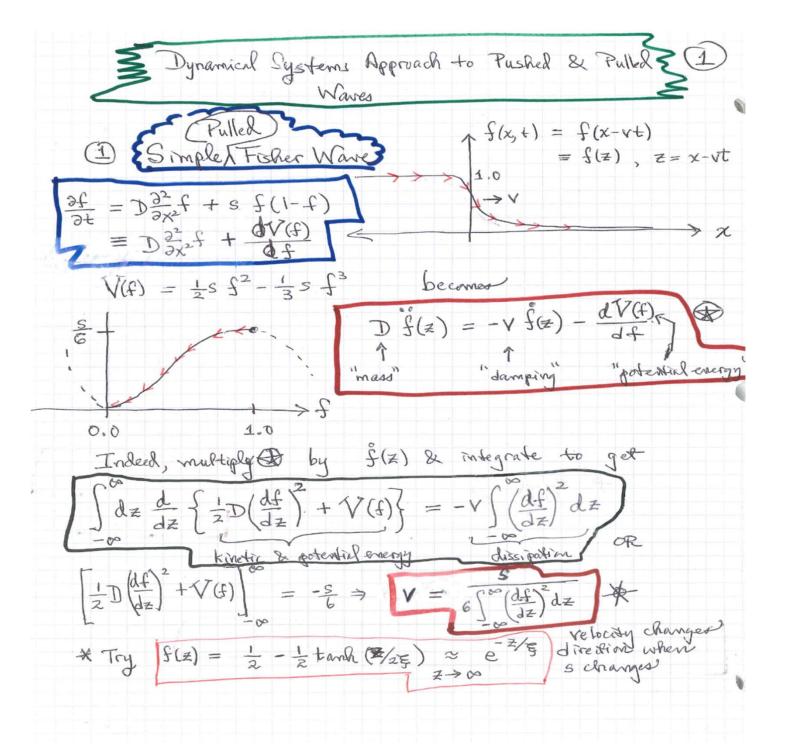
A convenient reparameterization

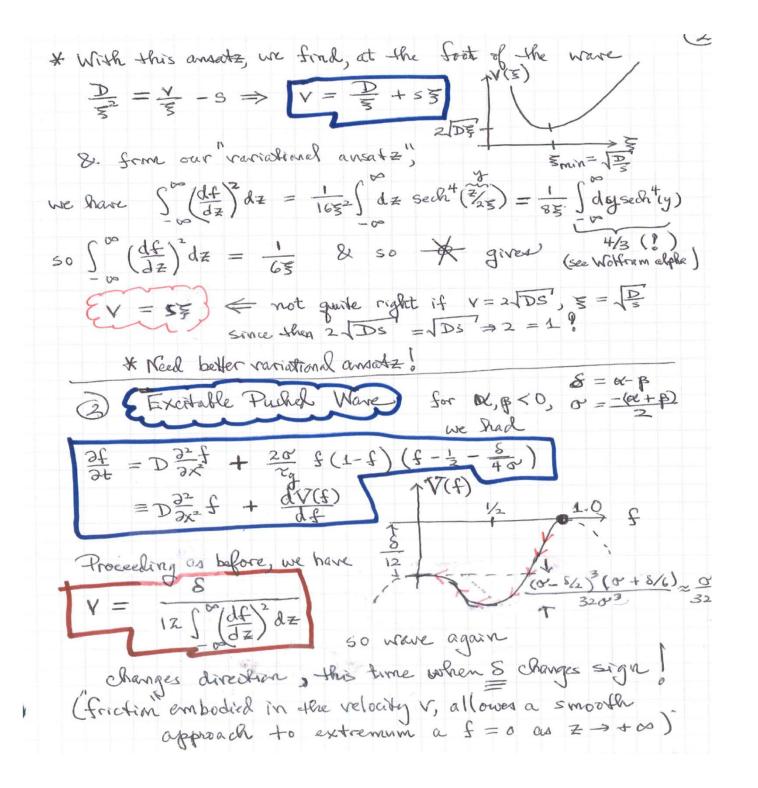
To study the antagonistic interactions in the third quadrant, we rotate our coordinates to $\sigma = -(\alpha + \beta)/2$ and $\delta = \alpha - \beta$:

$$\partial_t f = D_s \nabla^2 f + f(1 - f) \left[\sigma(2f - 1) + \frac{\delta}{2} \right] + \sqrt{D_g f(1 - f)} \xi \qquad (1)$$

 $\delta = \alpha - \beta$ = selective advantage; $\sigma = (\alpha + \beta)/2$ = "antagonism"







Double well potential, nucleation theory and Maxwell construction

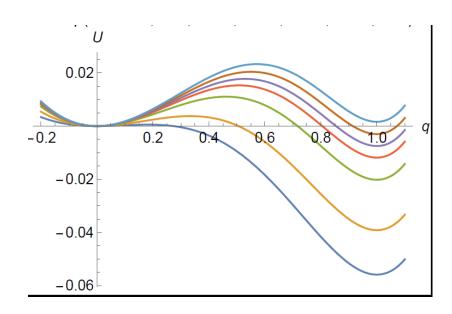
Recast dynamics in terms of a functional derivative:

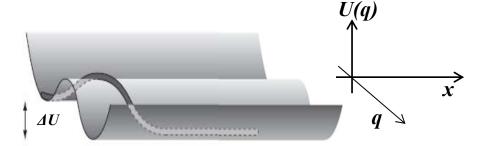
$$U(f) = -\frac{1}{\tau_g} \int_{0}^{q} 2\sigma f(1-f)(f-f^*) dq$$

$$\frac{\partial f(x,t)}{\partial t} = -\frac{\delta F}{\delta f(x,t)}, \quad F(t) = \int_{-\infty}^{\infty} \left[\frac{1}{2} \tau_g D \left(\frac{\partial f(y,t)}{\partial y} \right)^2 + U[f(y,t)] \right] dy$$

Can show that the dynamics is such that F(t) always decreases:

$$\frac{dF(t)}{dt} = -\int_{-\infty}^{\infty} \left(\frac{\partial f(x,t)}{\partial t}\right)^2 dx < 0 \quad [U(q) \text{ has a double well structure...}]$$





Pushed waves requiring a critical nucleus stall out when the two minima have the same depth

Pushed Fisher waves for α , β < 0

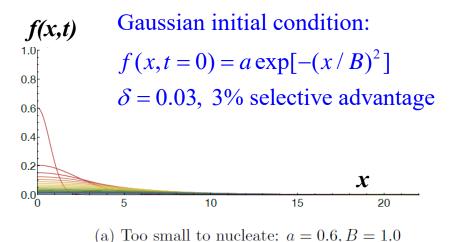
$$\frac{\partial f(x,t)}{\partial t} \approx D\nabla^2 f + \left(2\sigma/\tau_g\right) f(1-f)(f-f^*); f^* = \frac{1}{2} - \frac{\delta}{4\sigma},$$

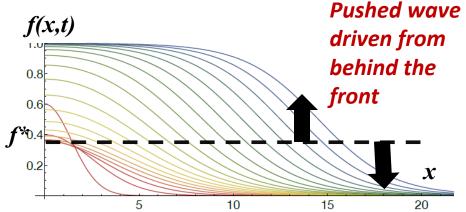
N. H. Barton & M. Turelli, The American Naturalist 178, E48 (2011)

an exact traveling wave solution exists!

$$f(x,t) = \frac{1}{1 + \exp[(x - v_E t)\sqrt{\sigma/\tau_g D}]}; \quad v_E = \frac{1}{2} \frac{\delta}{\sigma} \sqrt{D/\tau_g}; \quad \delta << \sigma$$

*A "critical propagule" size for the gene drive is required to nucleate the wave *Wave reverses direction when $\delta = \alpha - \beta$ changes sign





(b) Successful nucleation: a = 0.6, B = 2.0

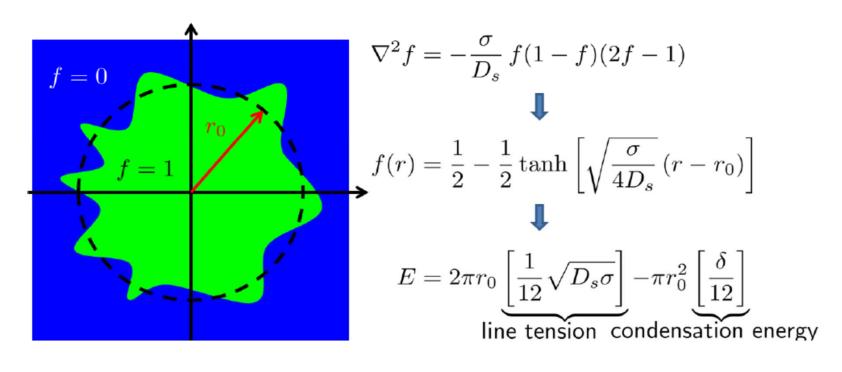
Nucleation has an even more dramatic effect in two dimensions

Nucleation and growth for $\delta > 0$, $\sigma \geq 0$

Suppose we look at droplets with $0 < \delta \ll \sigma$. The "energy" of such a drop is

$$E[f] = \int d\mathbf{x} \left[\frac{D_s}{2} (\nabla f)^2 + U(f) \right]$$

We can look for minimal E[f] solutions that look like droplets:



Test of nucleation theory in two dimensions

Xiaojue Zhu, R. Benzi, F. Toschi & drn

droplet energy $E(R) = 2\pi R \gamma - \pi c R^2$, line tension $\gamma = (2/3)\sqrt{\sigma/D}$, condensation energy $c = \delta/3D$ critical droplet size $= R_c = \gamma/c = (2/\delta)\sqrt{D\sigma}$

The dynamics of the droplet radius R(t) is given by

$$\frac{dR(t)}{dt} = -\frac{D}{R(t)} + \frac{\delta}{2} \sqrt{\frac{D}{\sigma}} \quad \text{(require } R(t) >> w = \text{ interface width)}$$

- \rightarrow critical droplet radius $R_c = \gamma / c = (2/\delta) \sqrt{D\sigma}$
- → dying droplets should vanish with a square root singularity,

$$R(t) = \sqrt{R_0^2 - 2D(t - t_0)},$$

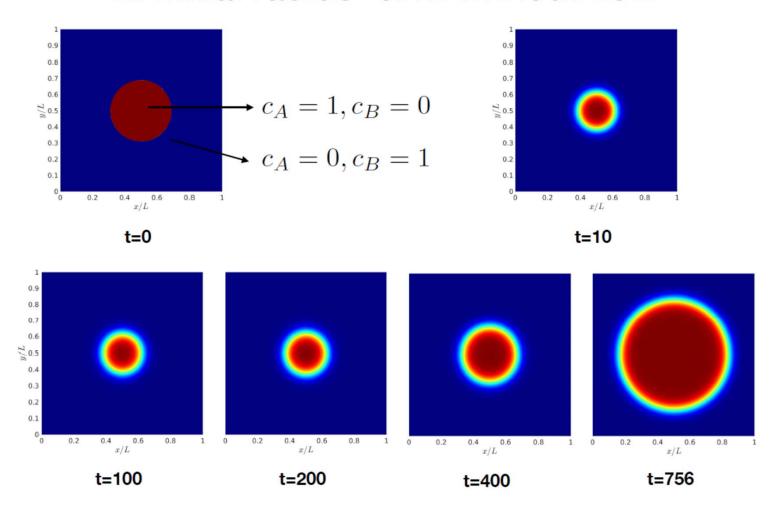
where R_0 is the radius of a dying droplet has well below the maximum R_c at time t_0

→ Once the droplet is above the maximum, we should eventually have a circular,

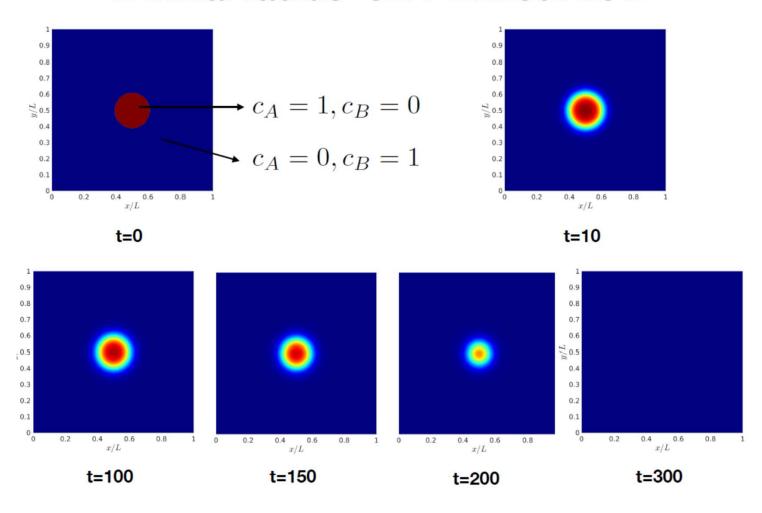
expanding pushed wave with
$$R(t) \approx vt$$
, $v = (\delta/2)\sqrt{D/\sigma}$

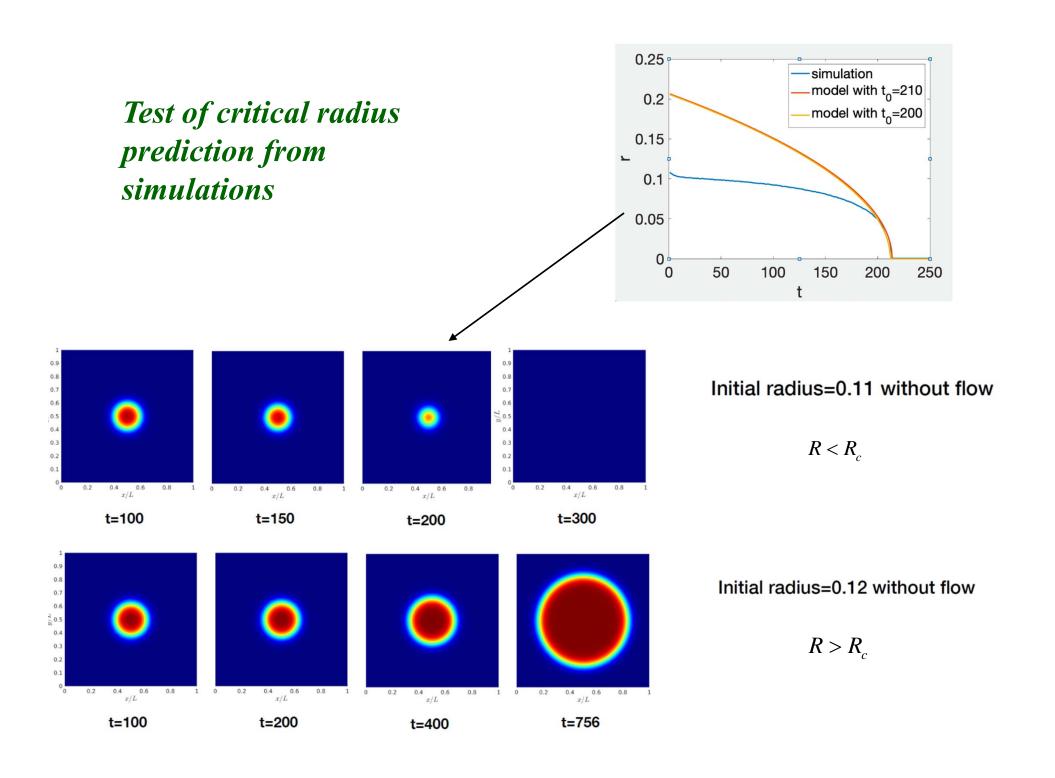
simulations: selective advantage = $\delta = \varepsilon_A - \varepsilon_B = 0.1$ antagonism = $\sigma = -(\varepsilon_A + \varepsilon_B)/2 = 0.25$

2. Initial radius=0.12 without flow



1. Initial radius=0.11 without flow

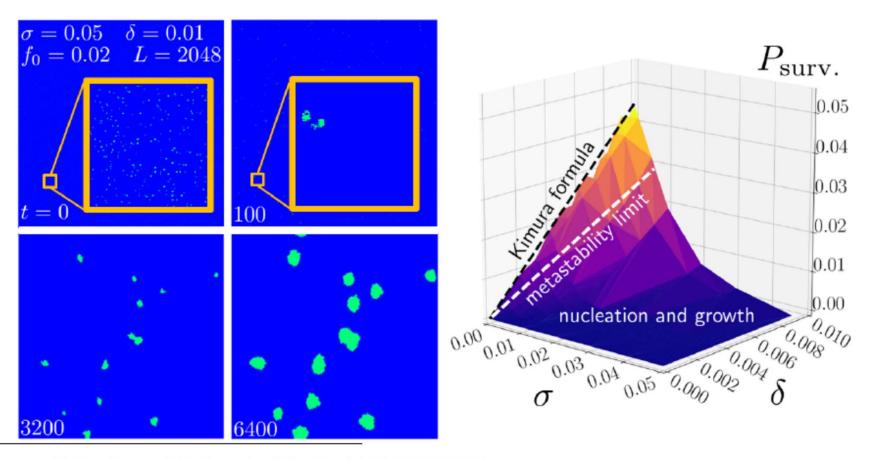




Green cells must overcome line tension to grow

$$\sigma = -(\alpha + \beta)/2$$
, $\delta = \alpha - \beta$, $\delta \ll \sigma \ll 1$

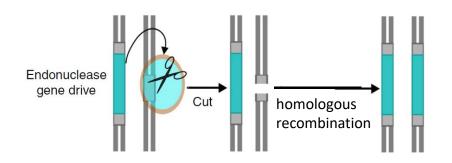
Nucleation theory ideas can be used to compute the survival probability in this limit



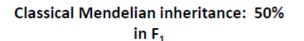
C. R. Doering, C. Mueller, and P. Smereka, Physica A 325, 243 (2003)

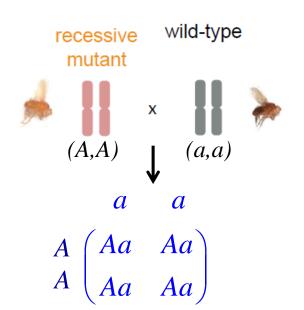
T. Maruyama, Theor. Popul. Biol. 5, 148 (1974)

Relevance to CRISPR-Cas9 gene drives & non-Mendelian population genetics

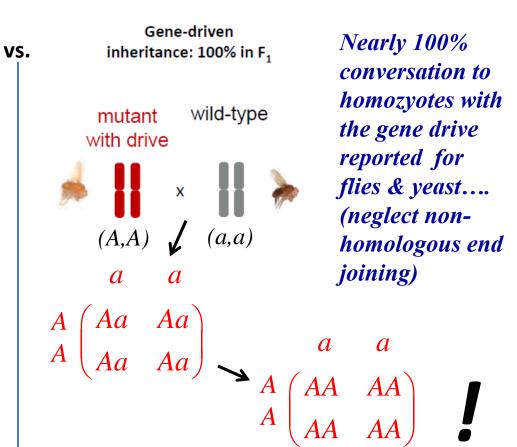


R. L. Unckless, A. G. Clark, and P. W. Messer, Genetics 205.2 (2017): 827-841.

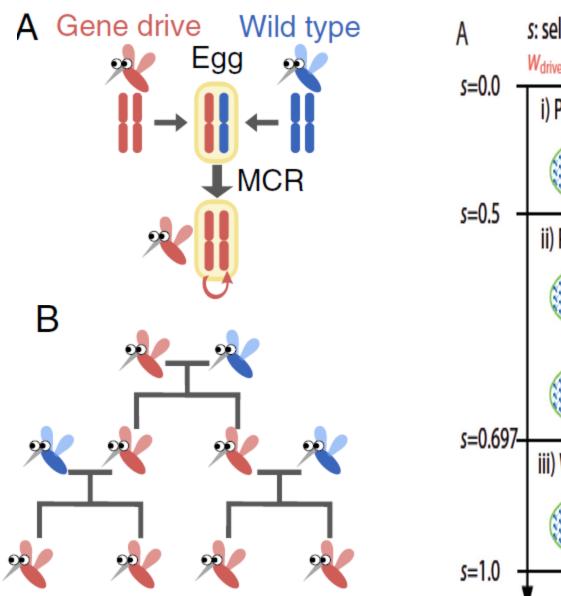


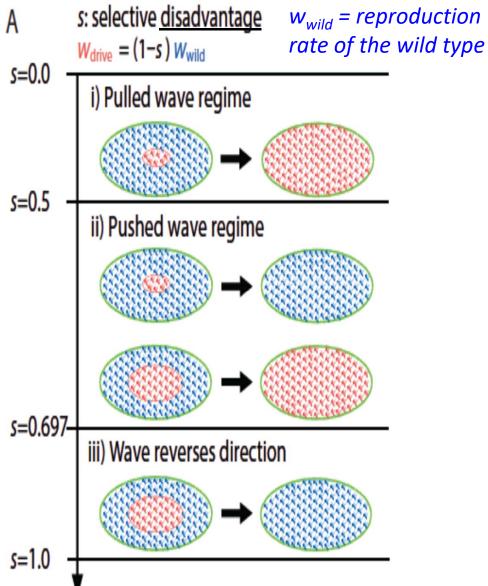


The offspring are heterozygous, and they display the wild type phenotype if dominant



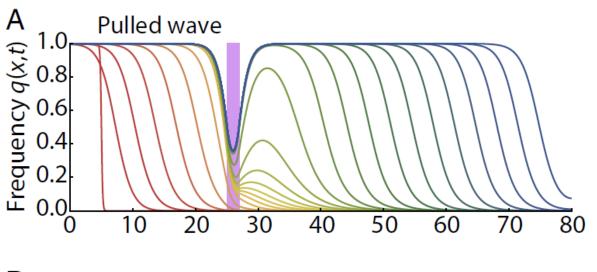
Spatial spread of pushed and pulled genetic waves

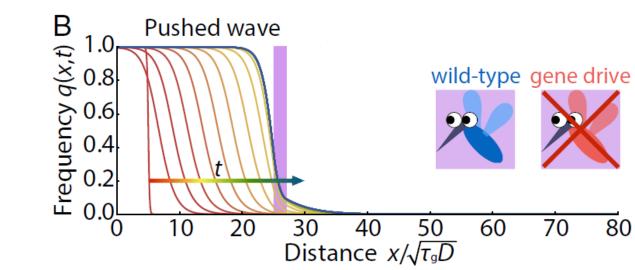




The effect of obstacles: Pushed, excitable waves are more fragile than their pulled Fisher wave counterparts...

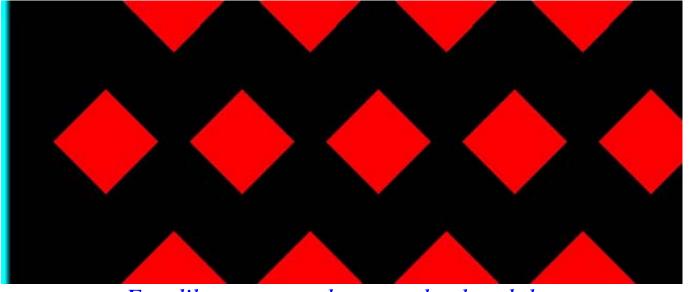
Stopping power of a selective disadvantage barrier in one dimension





In two dimensions, conventional Fisher waves easily traverse an array of obstacles

Simulations by W. Moebius, drn & A. Murray



For dilute arrays, the cusps heal and the propagation velocity is unchanged

But pushed waves are more fragile!

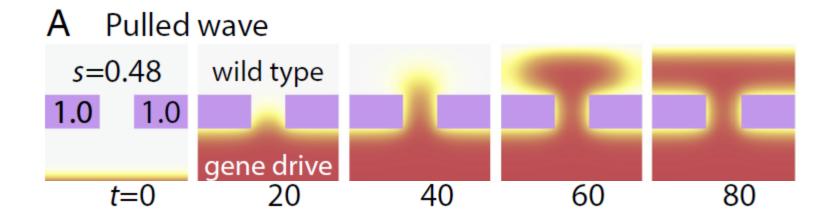


A dense array of obstacles merely leads to a reduced propagation velocity --

Stopping power of selective disadvantage barriers in two dimensions

$$\frac{\partial f(\vec{x},t)}{\partial t} \approx D\nabla^2 f + \left(2\sigma/\tau_g\right) f(1-f)(f-f^*) + \text{genetic drift}, \quad f^* = 1/2 + \delta/4\sigma$$

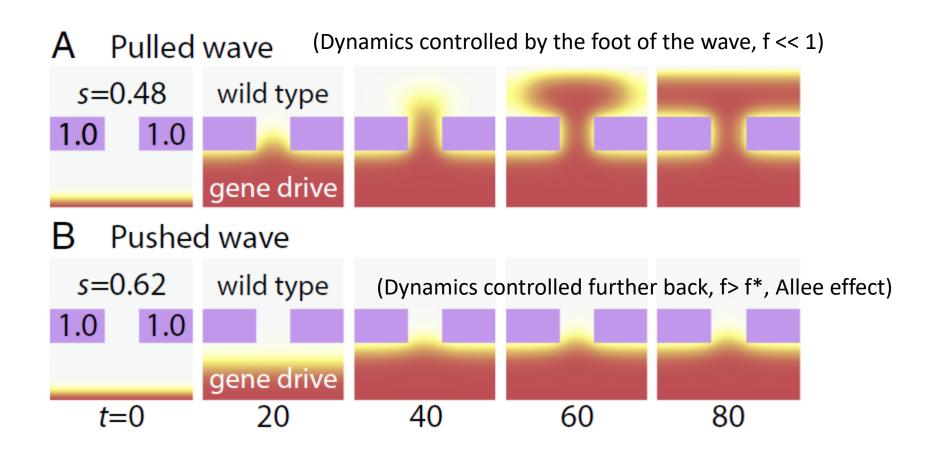
$$f(\vec{x},t) = \text{concentration of gene drive homozygotes at position } \vec{x} \text{ and time } t$$



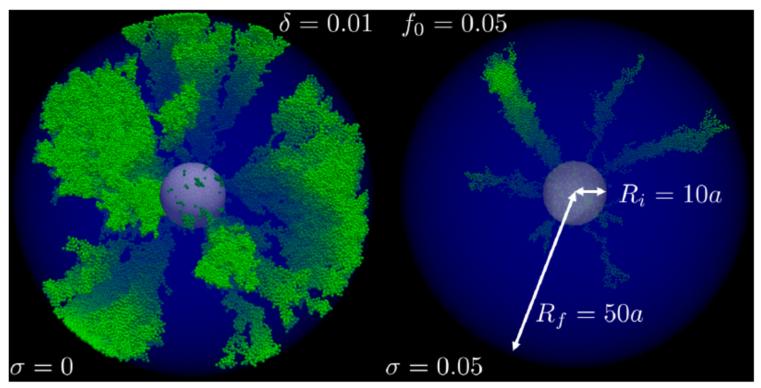
Stopping power of selective disadvantage barriers in two dimensions

$$\frac{\partial f(\vec{x},t)}{\partial t} \approx D\nabla^2 f + \left(2\sigma/\tau_g\right) f(1-f)(f-f^*) + \text{ genetic drift}$$

$$f(\vec{x},t) = \text{ concentration of gene drive homozygotes at position } \vec{x} \text{ and time } t$$



Nucleation in a growing spherical cluster



In a growing cluster of cells with inflating radius $R(t) = \alpha(t)R_0$ and $\alpha(t) = 1 + t/t^*$, the survival probability will be enhanced. One can see from the droplet dynamics $(\gamma = \sqrt{D_s \sigma}/12, c = \delta/12)$:

$$E = 2\gamma \alpha(t)\pi \bar{r}_0 - c[\alpha(t)]^2 \pi \bar{r}_0^2 \quad \Rightarrow \quad \bar{r}_0^* = \left[\int_0^\infty \frac{2\pi c t^* e^{-z} \, \mathrm{d}z}{z + 2\pi c t^*} \right] r_0^* \ll r_0^*$$

Pushed Genetic Waves and Antagonistic Interactions

- For pushed genetic waves, a "critical nucleus" is required to excite the wave to get it started (like nucleation theory)
- How can an excitable pushed wave from a gene drive be stopped? (They are fragile and can be stopped by obstacles....)
- Can we detect the existence of a critical nucleus with killer yeast or bacteria strains? (project with Andrea & Andrew)



Andrea



Andrew



Max Lavrentovich Univ. of Tennessee

Thank you!



http://streetanatomy.com